



extradurally. The arms of the clip were 1 mm wide, 0.5 mm thick and 8 mm long were curved to facilitate extradural placement around the cord. Compression injury was produced at the level of T1 by placing the clip extradurally for 30 seconds. The traumatised segment (S1) and adjoining upper segment of 1 mm length (S2) were removed for biochemical estimations. Sham operated controls had laminectomy done from C7 to T2 and the corresponding spinal segments were removed after one week from these rats. Immediately after surgery all rats received gentamycin 0.2 mg/100 g body weight (IM). The urinary bladders of all operated animals were manually compressed every 12 hours for one week following surgery. Water and food were made available *ad libitum*.

Dexamethasone and verapamil were administered daily for 7 days following injury. Dexamethasone was given in doses of 0.5 mg/kg on days one and 2, 0.25 mg/kg on days 3 and 4; 0.125 mg on days 5, 6 and 7. Verapamil was administered in doses of 0.2 mg/kg on days 1, 2 and 3; 0.4 mg/kg on days 4 and 5; 0.8 mg/kg on days 6 and 7. Control injured rats received saline.

The animals were evaluated for motor function for one week. Motor function was carried out on an inclined plane once in 2 days for 7 days. A blind testing method was used whereby the observer was not aware whether a given animal was control or a treated animal. The inclined plane consisted of a adjustable plank covered by a rubber mat constructed after the apparatus described by Rivlin and Taylor (11). The angle of the plane was increased from 0° until the rat could not maintain its position for 5 sec. Rats were placed on the inclined plane and the maximum angle at which the rat could maintain its position for atleast 5 sec without falling was measured.

**Biochemical studies :** Acetylcholinesterase (AChE) and phospholipids were estimated in spinal cord segments. Phospholipids were estimated 24 hrs after injury without any drug administration. AChE was estimated in spinal

segments from sham controls and drug treated rats one week after injury.

**AChE :** Spinal segments were cleaned of blood, weighed and homogenates were used for enzyme assay. AChE was assayed by the method of Ellman et al (12) using acetylthiocholine as substrate. Butyl cholinesterase (BuChE) activity was also estimated using butyl thiocholine as substrate and in the presence of BW 284C51, a specific inhibitor of AChE to ascertain any activity due to BuChE. The specific activity of AChE was found to be inhibited (98%) when BW 284C51 was used as an enzyme inhibitor. This suggests that most of the activity is due to AChE. The specific activity of the enzyme was expressed as units of enzyme activity per mg protein. The protein was determined by the method of Lowry et al (13).

**Phospholipids :** The concentration of total and individual phospholipid in spinal cord segments were determined in chloroform - methanol extracts. Total lipid extracts were prepared by the method of Folch et al (14) by homogenizing the spinal cord tissues in 5 ml chloroform - methanol - 12 MHCl. Phospholipids were isolated by two dimensional thin layer chromatography (14). Separated phospholipids were visualised by exposure to iodine vapors and iodine positive spots were quantitated after acid digestion (15).

Statistical significance was tested by Students' t-test and a level of  $P < 0.05$  was considered as significant. Drug treated groups were compared with saline treated injured rats.

## RESULTS

The concentration of phospholipid is expressed as a function of wet weight of tissue. Total phospholipid phosphorus showed a significant decrease in the injured spinal cord segment at 24 hrs. Both phosphatidyl choline and phosphatidyl ethanolamine was found to be decreased in the injured segment (Table I).

AChE activity was decreased in the traumatised segment I after one week in the

TABLE 1 : Changes in phospholipid content in spinal cord injured rats.

Group	Total phospholipids	Phosphatidyl ethanolamine	Phosphatidyl choline
Controls	62.02±6.13 ***	16.56±0.52 **	12.43±1.24 *
Injured	40.17±2.76	09.91±1.50	07.82±1.00

P values less than \*0.05 \*\*0.01 \*\*\*0.02

Phospholipid values are expressed as nmol/mg wet weight. Number of rats in each group is 4.

injured rats. No significant change in the activity was observed in segment II. Dexamethasone in the given doses were found to reverse the decrease in AchE activity in the S1 segment of the injured rats (Fig. 1).

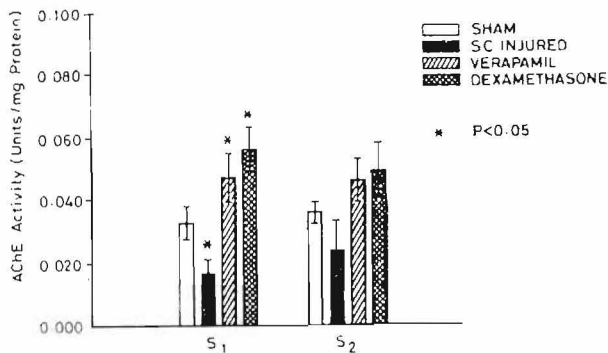


Fig. 1 : Changes in AchE activity in the spinal cord of rats S1, injured segment, S2 uninjured adjacent segment. Data are mean SEM of 4 rats in each group. Injured group is compared with sham controls, drug treated groups with the injured group.

Studies on neurologic function showed that sham operated rats could maintain a maximum angle of  $76.77 \pm 1.01$  in the inclined plane (Fig. 2). At one week assessment period, aneurysm clipped rats showed a decrease in the maximum angle in the inclined plane. The dexamethasone treated rats could maintain themselves at a higher angle than the injured rats at the end of one week. Verapamil treated rats also showed an improvement in the neurologic function. Neurologic recovery was better in the dexamethasone treated group ( $P < 0.001$ ).

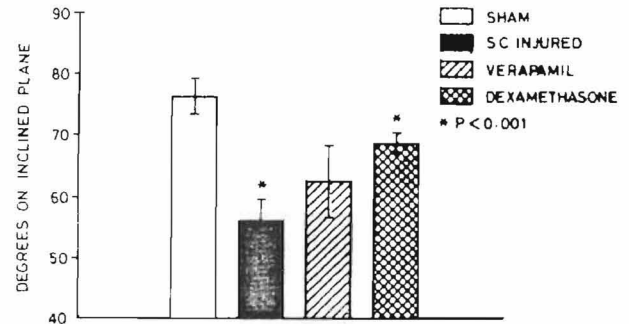


Fig. 2 : Motor function as observed on the inclined plane during one week after injury. Data are mean  $\pm$  SEM of 4 rats in each group. P values are indicated \*less than 0.001. Injured group is compared with sham controls. Drug treated groups are compared with the injured group.

## DISCUSSION

Results of this study show a decrease in phospholipid content in the injured tissue. Lipid hydrolysis and liberation of products of hydrolysis such as phospholipids and diacylglycerol are known to perturb the integrity of membranes (16). Prostaglandin levels are reported to be increased at the site of neuronal injury (17). Lipid peroxidation and generation of free radicals are some of the other biochemical events detected during the early phase of spinal cord injury (5, 6).

Change in membrane phospholipids may alter the activity of the enzymes which are functionally related to neuronal membrane. In the present study the activity of enzyme AchE is found to be decreased in the injured segment. Similar results have been observed in primates following spinal cord injury induced by weight drop (18). These changes are reversed by test drugs dexamethasone and verapamil. In the study on primates beneficial effects have been observed by these drugs on the various biochemical parameters such as AchE activity, NaKATPase and lysosomal enzymes during early phase of contusion injury. High dose of dexamethasone was reported to be effective in spinal cord compression (8). Weidefeld et al (19) has reported that dexamethasone is effective in lowering prostaglandin production in the brain.

The reported cerebroprotective mechanisms of methyl prednisolone include inhibition of post traumatic lipid peroxidation (20, 21) reversal of intracellular calcium accumulation (22) and prevention of neurofilament degradation (23).

Normal nerve cells maintain a remarkable calcium iongradient between intracellular and extracellular compartments. Ischaemic or traumatic insults to the neurones can alter this balance (24). These changes are attributed to the increase in cell membrane permeability and consequent influx of  $Ca^{2+}$ . Desphpande et al (25) have demonstrated that calcium accumulates before cell necrosis occurs implying that change in calcium homeostasis is an early event in neuronal injury. Accumulation of calcium in the intracellular space is considered as a factor activating phospholipase A and C, which will attack membrane phospholipids. Verapamil treatment for one week partially

reversed the changes in enzyme activity of injured segment and produced partial improvement in neuronal function. Verapamil may produce these effects by reducing the toxic effects of calcium accumulation and further damage to membranes.

In summary, this study highlights the biochemical changes observed during early phase of spinal cord injury induced by compression. The test drugs dexamethasone and verapamil reversed the change in acetylcholinesterase activity and improved neurologic function partially.

#### ACKNOWLEDGEMENTS

Authors wish to express thanks to J. Samuel, Swamidoss and Jesu for their technical assistance. Thanks are also due to Dr. W. R. Klemm, Texas, USA, for granting permission to do the study on phospholipids.

#### REFERENCES

1. Bingham WG, Goldman H, Friedman SJ et al. Blood flow in normal and injured monkey spinal cord. *Journal of Neurosurgery* 1975; 43 : 162-171.
2. Senter HJ, Venes JL. Altered blood flow and secondary injury in experimental spinal cord trauma. *Journal of Neurosurgery* 1978; 49 : 569-578.
3. Feldman RA, Yashon D, Locke GE, Hunt WE. Lactate accumulation in primate spinal cord during circulatory arrest. *Journal of Neurosurgery* 1971; 34 : 618.
4. Osterholm JL, Mathews GJ. A proposed biochemical mechanism for traumatic spinal cord hemorrhagic necrosis. Successful therapy for severe injuries by metabolic blockade. *Trans American Neurological Association* 1971; 96 : 187-191.
5. Demopoulos HB, Flamm ES, Pietronigro DD, Seligmann ML. The free radical pathology and the microvasculature in the major central nervous system disorders. *Acta Physiologica Scandinavica (suppl)* 1980; 492 : 91-119.
6. Anderson DK, Means ED. Iron induced lipid peroxidation in spinal cord, protection with mannitol and methyl prednisolone. *Journal of Free Radicals Biology and Medicine* 1982; 1 : 59-64.
7. Pryor WA, Stanley JP, Blair E. Auto oxidation of polyunsaturated fatty acids : II. A suggested mechanism for the formation of TBA - reactive material from prostaglandin like endoperoxides. *Lipids* 1976; 11 : 370-379.
8. Delattre JY, Arbit E, Thaler HT, Rosenblum MK, Posner JB. A dose-response study of dexamethasone in a model of spinal cord compression caused by epidural tumor. *Journal of Neurosurgery* 1989; 70 : 920-925.
9. Eidelberg EM, Staten E, Watkins AC, McGraw D, McFadden C. A model of spinal cord injury. *Surgical Neurology* 1976; 6 : 35-38.
10. Rivlin AS, Tatar CH. Regional spinal cord blood flow in rats after severe cord trauma. *Journal of Neurosurgery* 1978; 49 : 844-853.
11. Rivlin AS, Tatar CH. Objective clinical assessment of motor function after experimental spinal cord injury in the rat. *Journal of Neurosurgery* 1977; 47 : 577-581.
12. Ellman GL, Courtney KD, Andres V Jr, Featherstone K. A new and rapid calorimetric determination of acetylcholinesterase activity. *Biochem Pharmacol* 1961; 7 : 75-88.
13. Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with the folin phenol reagent. *Journal of Biological Chemistry* 1957; 193 : 265-275.

14. Falch, Less M, Stanley CHS. A simple method for the isolation and purification of total lipids from animal tissues. *Journal of Biological Chemistry* 1957; 226 : 497-509.
15. Bartlett RG. Phosphorous assay in column chromatography. *Journal of Biological Chemistry* 1959; 234 : 466-468.
16. Allan D, Thomas P, Michell RH. Rapid transbilayer diffusion of 1,2-dicetyl glycerol and its relevance to control of membrane curvature. *Nature (London)* 1978; 276 : 286-290.
17. Johnson HT Jr, Daniel HB. Altered levels of PGF in cat spinal cord tissue following traumatic injury. *Prostaglandins* 1976; 11 : 51-61.
18. Cherian L, Kuruvilla A, Abraham J. Changes in acetylcholinesterase during early phase of experimental spinal cord trauma in primates. *Indian Journal of Experimental Biology* 1989; 27 : 751-753.
19. Weidenfeld J, Sysy J, Shohami E. The effect of dexamethasone on prostaglandin synthesis in various brain areas of the rat. *Journal of Neurochemistry* 1987; 48 : 1351-1354.
20. Demopoulos HB, Flamm ES, Seligman ML, Pietrocigro DD, Tomasula J, DeCricito V. Further studies on free radical pathology in the major central nervous system disorders : Effect of very high doses of methyl prednisolone in the functional outcome, morphology and chemistry of experimental spinal cord impact injury. *Canadian Journal of Physiology and Pharmacology* 1982; 60 : 1415-1424.
21. Kurihara M. Role of monoamines in experimental spinal cord injury in rats. Relationship between Na+K+ATPase and lipid peroxidation. *Journal of Neurosurgery* 1985; 62 : 743-749.
22. Young W, Flamm ES. Effect of high dose of corticosteroid therapy on blood flow, evoked potentials and extra cellular calcium in experimental spinal injury. *Journal of Neurosurgery* 1982; 57 : 667-673.
23. Braughler JM, Hall ED. Effects of multidose methyl prednisolone sodium succinate administration on injured cat spinal cord neurofilament degradation and energy metabolism. *Journal of Neurosurgery* 1984; 61 : 290-295.
24. Schanne Fax, Kane AB, Young EE, Farber JL. Calcium dependence of toxic cell death : a final common pathway. *Science* 1979; 206 : 700-702.
25. Deshpande JK, Siesjo BK, Wieloch T. Calcium accumulation and neuronal damage in the rat hippocampus following cerebral ischaemia. *Journal of Cerebral Blood Flow and Metabolism* 1978; 7 : 89-95.